J Med Sci 2017;37(4):155-162 DOI: 10.4103/jmedsci.jmedsci 113 16

ORIGINAL ARTICLE



Effects of Isolation Rearing and Early Antipsychotic Intervention on Oxidative Stress-induced Apoptosis and Brain-derived Neurotrophic Factor in Hippocampus in a Rat Model of Schizophrenia

Szu-Nian Yang¹, Yu-Yin Yang², Fang-Jung Wan³, Chuen-Lin Huang⁴, Yia-Ping Liu^{2,3}

¹Department of Psychiatry, Taoyuan Armed Forces General Hospital, Taoyuan 32551, ²Department of Physiology and Biophysics, National Defense Medical Center, ³Department of Psychiatry, Tri-Service General Hospital, ⁴Medical Research Center, Cardinal Tien Hospital, Taipei, Taiwan, Republic of China

Background: Oxidative stress-induced neuronal dysfunction has been considered an essential factor for the development of schizophrenia. However, a longitudinal and causal relation between the impacts of developmental stress and oxidative stress remains unsolved. The present study aimed to examine whether the oxidative stress-relevant dysfunctions of the apoptotic index can be induced in rats of isolation rearing (IR, a rodent model of schizophrenia) and to see if the intervention of antipsychotics can reverse these dysfunctions. Materials and Methods: Pharmacological manipulation (risperidone [RIS] [1 mg/kg/day], olanzapine [OLA] [2.5 mg/kg/day], or saline [SAL] vehicle) was introduced 4 weeks (adolescence) or 8 weeks (young adulthood) after IR (i.e., rats were 7- or 11-week-old). The regime of RIS, OLA, or SAL was continued for 9 weeks. Locomotor activity was employed to validate the IR effect. Rats' hippocampus immediately after sacrifice was removed to measure messenger RNA expression of Bax, Bcl-2, brain-derived neurotrophic factor (BDNF) and the plasma level of nitric oxide (NO). Results: The results showed: (i) IR rats were more hyperactive. (ii) RIS may exert anti-apoptotic effects on IR rats, particularly at their adolescent age (as indexed by increased Bcl-2 and decreased Bax/Bcl-2 ratio). (iii) The therapeutic potential of RIS can be also observed in the change of BDNF in an age-independent manner, in which RIS effectively increased the BDNF level in IR but not social (SOC) rats. (iv) Plasma NO was not altered. Conclusion: The study results support the utility of the IR paradigm in exploring mental disorders with neurodevelopmental origin in which early pharmacological intervention may provide a therapeutic benefit in the overloaded oxidative stress and the dysfunction of BDNF.

Key words: Brain-derived neurotrophic factor, early antipsychotic intervention, isolation rearing, oxidative stress, schizophrenia

INTRODUCTION

Schizophrenia is a multidimensional mental disorder in which patients suffer from progressively psychosocial impairment and cognitive decline, including distortions of thought and perception (positive symptoms) and lack of motivation or spirit (negative symptoms) across time in a deteriorating manner.¹ Although the pathoetiology of this disorder remains unclear, oxidative stress-induced neuronal dysfunction has been considered a key factor.^{2,3}

Oxidative stress indicates an imbalance between the systemic manifestation of reactive oxygen species (ROS) and

Received: November 06, 2016; Revised: May 24, 2017; Accepted: April 17, 2017

Corresponding Author: Prof. Yia-Ping Liu, Department of Physiology and Biophysics, National Defense Medical Center, 161, Minchuan East Road, Section 6, Taipei, Taiwan 114, Republic of China. Tel: +8862 87923100 ext. 18614; Fax: +8862 87923153. E-mail: yiaping@ndmctsgh.edu.tw

the antioxidant defenses (i.e., an ability of the biological system to repair the damage caused by ROS).⁴ Excessive oxidative stress may finally lead to cellular apoptosis which is regulated by, for example, the Bcl-2 family in which Bcl-2 is anti-apoptotic but Bax is pro-apoptotic.⁵ Recently, increasing evidence suggests a relevance of oxidative stress and mental illness with long-term environmental or social distress impact.⁶ This is important because it exemplifies the possibility that life stress can exert its effects on the events at the cellular level.

This is an open access article distributed under the terms of the Creative Commons Attribution-NonCommercial-ShareAlike 3.0 License, which allows others to remix, tweak, and build upon the work non-commercially, as long as the author is credited and the new creations are licensed under the identical terms.

For reprints contact: reprints@medknow.com

How to cite this article: Yang SN, Yang YY, Wan FJ, Huang CL, Liu YP. Effects of isolation rearing and early antipsychotic intervention on oxidative stress-induced apoptosis and brain-derived neurotrophic factor in hippocampus in a rat model of schizophrenia. J Med Sci 2017;37:155-62.

On the other hand, brain-derived neurotrophic factor (BDNF) is a neurotrophic factor abundant in brain, responsible for regulating the events of neurotransmission, such as differentiation, growth, and plasticity. Its role in the pathogenesis of schizophrenia has been addressed recently in terms of the interplay with oxidative stress, however, elaboration on this hypothesis by performing studies with large sample sized in humans appears difficult due to ethical reasons, for example, it is difficult to separate these effects from the influence of antipsychotics because patients are seldom drug naive. A longitudinal and causal relation between the impacts of developmental stress and antipsychotics thus need to be clarified by using an appropriate animal model.

Rat of isolation rearing (IR) is a commonly used method in which weanling rats are increased in social separation. Generally, they are allowed to see, smell, and hear other rats but not to have social contact with them. The IR rats exhibit profound psychological, behavioral, and neurochemical changes when they grow up,9 similar to the full-blown symptoms of schizophrenia in patients' adulthood. IR rats are thus suitable for modeling mental disorders with a pathoetiology based on developmental hypothesis, 10 such as schizophrenia. The hallmark features of IR-induced behavioral changes are locomotor hyperactivity and a defect in sensorimotor gating function and are is particularly useful in examining the pathogenic/therapeutic mechanism of schizophrenia. 11,12 Here, if the neurodevelopmental process has a crucial impact on the oxidative stress-related abnormalities in schizophrenia, it is expected that these abnormalities may be revealed in rats reared in social isolation (i.e., IR rats), and pharmacological interventions at an early stage of IR might reverse the dysfunctions. For the pharmacological interventions, the treatment of schizophrenia has been greatly improved with the extensive use of second-generation antipsychotics (SGAs) in recent years. It helps schizophrenic patients by relieving both their positive and negative symptoms with a lower risk of extrapyramidal side effects.¹³

In the present study, we examined the IR effects on oxidative stress-relevant dysfunctions of apoptotic index and BDNF, and to see if clinical antipsychotics may reverse these dysfunctions. For these, we employed longitudinal regimes of two commonly used SGAs, risperidone (RIS) and olanzapine (OLA). IR rats in adolescence (7-week-old) or young adulthood (11-week-old) were randomly entered into a drug protocol (RIS [1 mg/kg/day] or OLA [2.5 mg/kg/day] for 9 weeks) to examine the IR and drug effects on the Bcl-2 and Bax and the BDNF in hippocampus. Hippocampal neurogenesis has recently been considered to play an important role in the development of schizophrenia, particularly in terms of receiving a stressful or traumatic impact early in life, 14-16 such as the long-term environmentally stressful situation in rats of IR. 17 Finally, as nitric oxide (NO)

may act in a positive feedback loop with BDNF to regulate synaptic plasticity, ¹⁸ but in an opposite manner as antioxidants do (schizophrenia is accompanied by increased NO but decreased antioxidant level), ¹⁹ the plasma level of NO would be measured in the present study too.

The result of the present study may help clarify the role of oxidative stress in IR-induced abnormalities and is of great relevance to the evidence-based clinical strategy that detecting and intervening in schizophrenia at an early stage may protect neurons from further deterioration and may lead to a better long-term outcome.²⁰⁻²²

MATERIALS AND METHODS

Animals

Male Sprague-Dawley rats (BioLASCO, Taiwan) were used in all experiments. The rats were 21–23-day-old and had been weaned on arrival at the animal center of the National Defense Medical Center (NDMC, Taipei, Taiwan). Rats in the IR group were housed singly. They could see, hear, and smell others but were kept separate and denied physical contact. The control group for IR was a social rearing (SOC) group. These rats were from the same batch as the IR rats but were reared socially (n = 2 for each cage). All rats were housed in a temperature (21°C-25°C) and humidity (40%-60%) controlled holding facility with 12 h light/dark cycles (light on from 700 to 1900) and received food and water ad libitum. After weaning (3 weeks postnatal age), rats were randomly and equally assigned to the IR or SOC group. Pharmacological manipulation (RIS [1 mg/kg/day], OLA [2.5 mg/kg/day], or saline [SAL] vehicle) was introduced 4 weeks (adolescence) or 8 weeks (young adulthood) later (i.e., rats were 7- or 11-week-old). The regime of RIS, OLA, or SAL was continued for 9 weeks. Locomotor activity was employed to validate the IR effect. Immediately after completion of the final blood sampling, the rats were sacrificed, and their hippocampus was removed to measure messenger RNA (mRNA) expression of Bax, Bcl-2, and BDNF. All experimental procedures were approved by the Laboratory Animal Center from the NDMC (AAALAC full accreditation).

Locomotor activity

Locomotor activity was measured using a computerized automated activity monitoring system (MED Associates, Inc., St. Albans, VT, USA). The system included four plexiglass chambers (43 cm \times 43 cm \times 30 cm) equipped with an infrared array of 16 photodetectors and corresponding light sources that emitted photobeams 3 cm apart and 4.5 cm above the chamber floor. Travel distance was recorded at the assigned intervals and analyzed with Med-Associates software.

Szu-Nian Yang, et al.

Messenger RNA expression of hippocampal Bax, Bcl-2, and brain-derived neurotrophic factor

Total RNA of rat hippocampus was extracted by according to manufacturer's protocol of EZ-RNA II total RNA isolation kit (Catalog No. 20-410-100, Biological Industries, USA). The quality and concentration of total RNA were determined by spectrophotometry (NanoDrop ND-1000® Spectrophotometer, Thermo Scientific). Subsequently, one microgram of total RNA was reverse-transcribed into complementary DNA using the RNA-to-Ct[™] 1-Step Kit (One-Step Real-Time-Polymerase chain reaction [RT-PCR] Master Mixes, Thermo Scientific). Specific primers for rat Bax (Rn01480160 g1), Bcl-2 (Rn99999125 m1), **BDNF** (Rn02531967 s1) GAPDH (Rn99999916 s1) were purchased from Applied Biosystems (USA) and to use for quantitative RT-PCR (7500 Real-Time PCR System, Applied Biosystems, USA). The reaction was incubated at 50°C for 2 min, 95°C for 10 min, followed by forty cycles of denature at 95°C for 15 s and annealing/extension at 60°C for 30 s. All mRNA quantities of target genes were analyzed in triplicates. GAPDH gene expression was used as an internal control standard for normalization and the SAL-treated group served as a calibrator sample for the comparative $2^{\Delta\Delta Ct}$ method.^{23,24}

Nitric oxide

NO production was indexed by the amount of nitrite and nitrate. Plasma NO was measured in terms of nitrate/nitrite levels following adopted from methods of Green et al.²⁵ and Kalaivani et al.²⁶ In brief, plasma, SAL and 5-sulfosalicylic acid were added. The precipitate was removed by centrifugation at 10,000 rpm for 10 min, 4°C. Sample (80 µl), enzyme cofactor mixture (10 µl), and nitrate reductase mixture (10 µl) were added by sequence and placed for 3 h at room temperature. Griess reagents (50 µl R1 and 50 µl R2 were added by sequence). The mixture was then allowed to stand for 20 min. The absorption at 540 nm was measured in spectrophotometer (Thermo Scientific Multiskan, Scan Jose, CA, USA).

Drug

RIS and OLA (Sigma-Aldrich Co., LLC) were dissolved in 1 N acetic acid and titrated to a final pH of 7 with 1 N NaOH. Drugs were prepared to produce a total injection volume of 1.0 ml/kg and were injected subcutaneously for 9 weeks (RIS 1 mg/kg/day, OLA 2.5 mg/kg/day and SAL at the same injection volume). The dosages used in this study have previously been shown to recover prepulse inhibition (PPI) in rats.^{27,28}

Statistical analyses

Statistical analyses were performed across the groups using a multi-factor analysis of variance using rearing condition, treatment, and timing as the between-subjects factor. Further analyses and *post hoc* multiple comparisons were performed if necessary. For brevity, F-values are only provided for significant effects. The value of P < 0.05 was considered statistically significant.

RESULTS

Locomotor activity

For locomotor activity at baseline, IR/older rats were more hyperactive. There were main effects of rearing condition ($F_{(1,60)}=23.12,\,P=0.000$) and intervention age ($F_{(1,60)}=18.80,\,P=0.000$) (adolescent IR: 7145 ± 402; adolescent SOC: 5710 ± 329; young adult IR: 9790 ± 442; young adult SOC: 7237 ± 385). Treatment with RIS or OLA had no effect on locomotor activity.

Bax, Bcl-2

Several indexes of apoptosis in the hippocampus were investigated with PCR. Our results revealed that for the SAL regime, lower Bax levels were found in adolescent rats (intervention age, $F_{(1,20)}=5.58,\ P<0.05$) which were due to a higher Bax level in young adult SOC rats. A higher Bcl-2 level was observed in rats treated with RIS (treatment, $F_{(1,40)}=8.85,\ P<0.01$), which was due to the higher Bax levels in both adolescent and young adult IR rats. Our results also showed that RIS but not OLA reduced the ratio of Bax/Bcl-2 in IR-adolescent rats (main effect of rearing condition, $F_{(1,20)}=5.14,\ P<0.05,\$ and effect of rearing condition × intervention age, $F_{(1,20)}=4.76,\ P<0.05$). Post hoc multiple comparisons confirmed that it was due to the difference between RIS/adolescent and SAL/adolescent rats [Figures 1 and 2].

Brain-derived neurotrophic factor

Our results showed that RIS and OLA can increase the BDNF level (treatment, $F_{(1,40)} = 12.6$, P < 0.01). RIS and OLA-treated IR rats had an overall higher BDNF level ($t_{(22)} = 3.84$, P < 0.01 for RIS and $t_{(22)} = 2.16$, P < 0.05 for OLA) [Figure 3].

Nitric oxide

There were no statistically significant effects of rearing condition, intervention age, and treatment in amount of plasma NO [Figure 4].

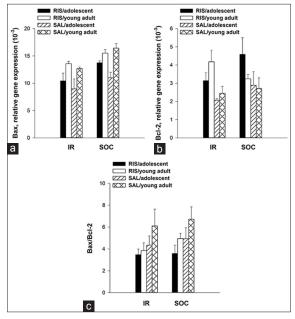


Figure 1: For saline regime, lower Bax levels appear in the adolescent rats which were due to a higher Bax level in young adult SOC rats (a). Risperidone-treated isolation rearing rats (both adolescent and young adult rats) have a higher Bcl-2 level (b). Risperidone also reduces the ratio of Bax/Bcl-2 particularly in adolescent isolation rearing rats (c). For each subgroup, n = 6. Data are represented as group average + standard error of mean. (IR: Isolation reared rats; SOC: Social control rats; RIS: Risperidone; OLA: Olanzapine; SAL: Saline)

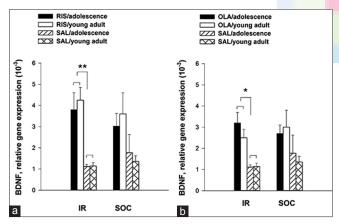


Figure 3: Risperidone (a) and olanzapine (b) can increase the brain-derived neurotrophic factor level. The drug-increased brain-derived neurotrophic factor is more pronounced in isolation rearing rats (pooled groups of different age, for each subgroup, n = 6). Data are represented as group average + standard error of mean. *P < 0.05 versus saline, *P < 0.01 versus saline. (IR: Isolation reared rats; SOC: Social control rats; RIS: Risperidone; OLA: Olanzapine; SAL: Saline)

DISCUSSION

The oxidative stress-induced dysfunctions are crucial in the pathoetiology of schizophrenia, and as the latter is a disorder with the developmental origin, an animal model of developmental impact on brain function appears suitable paradigm to examine

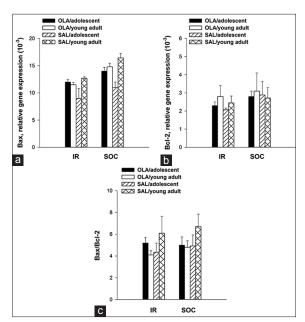


Figure 2: There are no statistically significant effects of rearing condition, intervention age and treatment in Bax level (a), Bcl-2 (b) and the ratio of Bax/Bcl-2 (c). For each subgroup, n = 6. Data are represented as group average + standard error of mean. (IR: Isolation reared rats; SOC: Social control rats; RIS: Risperidone; OLA: Olanzapine; SAL: Saline)

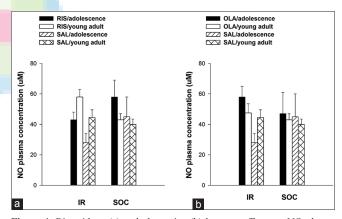


Figure 4: Risperidone (a) and olanzapine (b) have no effects on NO plasma concentration. There are no statistically significant effects of rearing condition, intervention age, and treatment in plasma nitric oxide. For each subgroup, n = 6. Data are represented as group average + standard error of mean. (IR: Isolation reared rats; SOC: Social control rats; RIS: Risperidone; OLA: Olanzapine; SAL: Saline)

the hypothesis that the neurodevelopmental process has a crucial impact on the oxidative stress-related abnormalities. The present study initially confirmed the validity of IR by showing IR-induced locomotor hyperactivity, and then for the first time demonstrated that RIS may exert anti-apoptotic effects on IR rats, particularly at their adolescent age (as indexed by increased Bcl-2 and decreased Bax/Bcl-2 ratio). The therapeutic potential of RIS can be also observed in the change of BDNF in an age-independent manner, in

which RIS effectively increased the BDNF level in IR but not SOC rats. Our results, in general, support the hypothesis that developmental impact may reflect in the oxidative stress level. These findings are further discussed as the following.

The present study demonstrated a varied sensitivities and impact of IR and SGAs acting on the oxidative stress-relevant biological variables. Whereas only RIS may be beneficial for younger generation (i.e., adolescent stage) of IR rats in terms of the anti-apoptotic change (as seen in the increased Bcl-2 level and decreased Bax/Bcl-2 ratio), both RIS and OLA were helpful for IR rats in an age-independent manner in terms of the neurogenesis (as seen in the high levels of BDNF in IR but not SOC rats, no matter when was the intervention timing). The underlying mechanism may be complicated. However, it could be interpreted in several aspects.

First, compared to BDNF, Bcl-2 family is a more direct pathway to reflect the imbalance of oxidative stress,4 the therapeutic effects of RIS appear more pronounced in younger generation of IR rats, suggesting a developmental specificity of IR-induced characteristics. Second, the antipsychotic effects on oxidative stress are controversial, while RIS previously has been reported to augment apoptosis through enhancing Bax in a stroke rodent model²⁹ and reducing Bcl-2/bax gene expression ratio in an in vitro study, 30 Keilhoff et al. on the contrary reported that RIS can promote the survival of young neurons in the hippocampus by enhancing the expression of the anti-apoptotic protein Bcl-2.31 Our hippocampal results apparently support the study of Keilhoff's group and highlight the specific feature of hippocampus, in which it is among the most important brain areas to interpret the pathoetiology of schizophrenia.32-34

BDNF, on the other hand, interpret the pathogenesis of schizophrenia from an aspect of neuronal function, including brain development, neuroplasticity, and synaptic connectivity.35 Cognitive dysfunction and epigenetic alterations of the BDNF gene are induced by social isolation during early adolescence.36 The involvement of BDNF in the IR is highly possible, as BDNF is involved in the chloride homeostasis-related central expression of KCC2/NKCC1,³⁷ which can be also adjusted by IR in SPAK knockout mice. 12 In terms of the relationship between oxidative stress and BDNF system, although interactions between them were reported, which is along with the developmental hypothesis, 8 the results of the present study indicate that the change of BDNF appears not the primary outcome following oxidative stress such as the Bcl-2 family. Further study to elucidate the causal relationships between oxidative stress and the BDNF system is necessary. The BDNF data are of particular relevance to research on schizophrenia, as the substance is active during a critical period of neuroplasticity in schizophrenia development. 38,39 However, the neuroprotective profile of antipsychotics varies. For example, BDNF gene expression is increased by olanzapine,⁴⁰ but decreased by haloperidol and a high dose of RIS.⁴¹ Our data demonstrated that chronic RIS administration at a relative low dosage (i.e., 1 mg/kg) might benefit BDNF gene expression, possibly through its 5HT, blockade activity.⁴¹

It is noted that in the present study, RIS seems superior to OLA in adjusting the balance of oxidative stress in younger IR rats, indicating a better therapeutic index of RIS in restoring the abnormalities with developmental origin, which is along with what reported previously that long-term RIS but not OLA to regulate the factors relevant to oxidative stress.⁴² Note in terms of medicinal chemistry, RIS belongs to the benzisoxazole class, which is different from OLA, a commonly used SGA belonging to thienobenzodiazepine class. Here, the superiority of RIS than OLA in restoring IR-induced abnormalities is a cross-model phenomenon as it was also reported in a paradigm of repeated treatment with phencyclidine, a substance known to induced PPI deficit in rodents and psychosis in human.^{43,44}

The duration of antipsychotic intervention could be one of the factors to determine its effects on oxidative stress. Previous evidence revealed that a shorter antipsychotic regime causes sabotage effects, however a longer one (for example, 180 days, rather than 90 days) turns to be beneficial in terms of the antioxidant enzymes and membrane lipid peroxidation. At As the drugged time in our study was 9 weeks, it possible explains the observation that the failure of RIS effects on Bax, as the intervention duration is insufficient to cause a valid effect.

Note although NO may serve as a signaling molecule involves in the development and treatment of schizophrenia, the alterations of NO metabolism are not unique to schizophrenia, thus NO is not a suitable biological index for this disease. 45 The involvement of NO in schizophrenia is complicated, as both overproduction and underproduction of NO have been suggested. 46,47 This study showed that the plasma NO was not sensitive to both IR procedure and antipsychotic regimes. It is also possible that the peripheral NO fails to reflect the pathology of schizophrenia in central nervous systems and the benefit bringing from the antipsychotics (i.e., enhancement of nitrergic activity). 47

It is interesting that in the present study, only IR rats appeared effective to the RIS treatment. This is important for clinical implication in terms of the concept of early antipsychotic intervention. In general, our results add to the evidence to support the concept of early antipsychotic intervention. ¹² Thus, how early to start antipsychotic treatment is debatable. For those who consider not taking antipsychotics in the prodromal phase of schizophrenia based on the issues of false-positive

diagnosis, 48,49 risking of stigmatizing patients, 50,51 and the long-term side effects of drugs, 52,53 and thus results in an ethical dilemma regarding early antipsychotic intervention.⁵⁴ On the other hand, if considering schizophrenia a developmental disorder as hypothesized, the biological method should help impede the progress of the disorder and prevent the patients from receiving too much biochemical disturbance before they are aware of the symptoms, thus the pharmacological interventions at an early stage of IR might be beneficial. The study apparently supports the concept of early antipsychotic intervention, in a more general viewpoint of stress. Thus, the present study demonstrates an example in which a long-term environmental stress may cause effects onto the cellular events of ROS-oxidative stress and the employment of antipsychotic regime in an earlier stage of IR may be beneficial. Further as RIS did not cause any negative effect on Bax/Bcl-2 or BDNF in adolescent rats compared to young adult controls, and this result may demonstrate a safety profile for early treatment in prodromal schizophrenic patients.

CLINICAL IMPLICATION AND CONCLUSION

Although pharmacological intervention with antipsychotics has been proven effective in managing the symptoms of schizophrenic patients, an earlier intervention is an ethical dilemma. False-positive diagnosis (as the symptoms are not full-blown), potential risk of stigmatizing patients, and the long-termside effects appear against the earlier pharmacological intervention. On the other hand, if considering schizophrenia to be a developmental disorder, the biological method should help impede the progress of the disorder thus prevent the patients from receiving too much biochemical disturbance before they are aware of the symptoms. ¹² The present study to a degree supports the early intervention of SGAs in terms of the oxidative stress-induced dysfunctions.

The hippocampal data obtained from the present study may support the use of SGA in managing the apoptosis-related abnormalities. In general, developmental stage plays a crucial role in determining the pharmacological efficacy onto the oxidative-stress relevant dysfunctions, thus earlier pharmacological intervention appears more beneficial in reversing the overloaded oxidative stress-induced abnormalities, as RIS may exert anti-apoptotic effects on IR rats at their adolescent age (as indexed by increased Bcl-2 and decreased Bax/Bcl-2 ratio), which is also in line with the sensorimotor gating profile. Our results support the neurodevelopmental hypothesis of schizophrenia and validate the utility of the IR paradigm in exploring mental disorders with neurodevelopmental origin. In terms of translational medicine, our results also demonstrated that the oxidative

stress profile can be one of the biological markers reflecting the efficacy of antipsychotics, in which earlier intervention would lead to a better progress.

Financial support and sponsorship

This research was supported by grant from Taoyuan Armed Forces General Hospital (105-12), National Science Council (NSC 96-2314-B-073-001) and National Defense Medical Center (MAB102-83) of Taiwan.

Conflicts of interest

There are no conflicts of interest.

REFERENCES

- Remschmidt H. Early-onset schizophrenia as a progressive-deteriorating developmental disorder: Evidence from child psychiatry. J Neural Transm (Vienna) 2002;109:101-17.
- GoffDC,RomeroK,PaulJ,MercedesPerez-RodriguezM, Crandall D, Potkin SG. Biomarkers for drug development in early psychosis: Current issues and promising directions. Eur Neuropsychopharmacol 2016;26:923-37.
- 3. Koga M, Serritella AV, Sawa A, Sedlak TW. Implications for reactive oxygen species in schizophrenia pathogenesis. Schizophr Res 2016;176:52-71.
- Dodson M, Darley-Usmar V, Zhang J. Cellular metabolic and autophagic pathways: Traffic control by redox signaling. Free Radic Biol Med 2013;63:207-21.
- Bazan NG. Lipid signaling in neural plasticity, brain repair, and neuroprotection. Mol Neurobiol 2005;32:89-103.
- Non AL, Binder AM, Kubzansky LD, Michels KB. Genome-wide DNA methylation in neonates exposed to maternal depression, anxiety, or SSRI medication during pregnancy. Epigenetics 2014;9:964-72.
- 7. Branchi I, Francia N, Alleva E. Epigenetic control of neurobehavioural plasticity: The role of neurotrophins. Behav Pharmacol 2004;15:353-62.
- 8. Zhang XY, Chen DC, Tan YL, Tan SP, Wang ZR, Yang FD, *et al.* The interplay between BDNF and oxidative stress in chronic schizophrenia. Psychoneuroendocrinology 2015;51:201-8.
- 9. Hall FS. Social deprivation of neonatal, adolescent, and adult rats has distinct neurochemical and behavioral consequences. Crit Rev Neurobiol 1998;12:129-62.
- Fone KC, Porkess MV. Behavioural and neurochemical effects of post-weaning social isolation in rodents-relevance to developmental neuropsychiatric disorders. Neurosci Biobehav Rev 2008;32:1087-102.

- Liu YP, Kao YC, Tung CS. Critical period exists in the effects of isolation rearing on sensorimotor gating function but not locomotor activity in rat. Prog Neuropsychopharmacol Biol Psychiatry 2011;35:1068-73.
- 12. Yang YY, Lu CL, Lo SM, Peng CH, Liu YP. Early antipsychotic intervention and schizophrenia. Med Hypotheses 2015;85:367-70.
- 13. Tandon R. Antipsychotics in the treatment of schizophrenia: An overview. J Clin Psychiatry 2011;72 Suppl 1:4-8.
- Humphrey WM, Dong H, Csernansky CA, Csernansky JG. Immediate and delayed hippocampal neuronal loss induced by kainic acid during early postnatal development in the rat. Brain Res Dev Brain Res 2002;137:1-12.
- Xu H, Qing H, Lu W, Keegan D, Richardson JS, Chlan-Fourney J, et al. Quetiapine attenuates the immobilization stress-induced decrease of brain-derived neurotrophic factor expression in rat hippocampus. Neurosci Lett 2002;321:65-8.
- 16. DeCarolis NA, Eisch AJ. Hippocampal neurogenesis as a target for the treatment of mental illness: A critical evaluation. Neuropharmacology 2010;58:884-93.
- 17. Lukkes JL, Burke AR, Zelin NS, Hale MW, Lowry CA. Post-weaning social isolation attenuates c-Fos expression in GABAergic interneurons in the basolateral amygdala of adult female rats. Physiol Behav 2012;107:719-25.
- 18. Cheng A, Wang S, Cai J, Rao MS, Mattson MP. Nitric oxide acts in a positive feedback loop with BDNF to regulate neural progenitor cell proliferation and differentiation in the mammalian brain. Dev Biol 2003;258:319-33.
- 19. Noto C, Ota VK, Gadelha A, Noto MN, Barbosa DS, Bonifácio KL, *et al.* Oxidative stress in drug naïve first episode psychosis and antioxidant effects of risperidone. J Psychiatr Res 2015;68:210-6.
- Linszen DH, Dingemans PM, Lenior ME, Scholte WF, de Haan L, Goldstein MJ. Early detection and intervention in schizophrenia. Int Clin Psychopharmacol 1998;13 Suppl 3:S31-4.
- 21. Perkins DO. Evaluating and treating the prodromal stage of schizophrenia. Curr Psychiatry Rep 2004;6:289-95.
- Keefe RS, Sweeney JA, Gu H, Hamer RM, Perkins DO, McEvoy JP, et al. Effects of olanzapine, quetiapine, and risperidone on neurocognitive function in early psychosis: A randomized, double-blind 52-week comparison. Am J Psychiatry 2007;164:1061-71.
- 23. Lameu C, Trujillo CA, Schwindt TT, Negraes PD, Pillat MM, Morais KL, *et al.* Interactions between the NO-citrulline cycle and brain-derived neurotrophic factor in differentiation of neural stem cells. J Biol

- Chem 2012;287:29690-701.
- 24. Livak KJ, Schmittgen TD. Analysis of relative gene expression data using real-time quantitative PCR and the 2(-Delta Delta C(T)) method. Methods 2001;25:402-8.
- Green LC, Wagner DA, Glogowski J, Skipper PL, Wishnok JS, Tannenbaum SR. Analysis of nitrate, nitrite, and [15N] nitrate in biological fluids. Anal Biochem 1982;126:131-8.
- 26. Kalaivani P, Saranya RB, Ramakrishnan G, Ranju V, Sathiya S, Gayathri V, et al. Cuminum cyminum, a dietary spice, attenuates hypertension via endothelial nitric oxide synthase and NO pathway in renovascular hypertensive rats. Clin Exp Hypertens 2013;35:534-42.
- Bakshi VP, Swerdlow NR, Braff DL, Geyer MA. Reversal of isolation rearing-induced deficits in prepulse inhibition by Seroquel and olanzapine. Biol Psychiatry 1998;43:436-45.
- 28. Zhang M, Ballard ME, Unger LV, Haupt A, Gross G, Decker MW, *et al.* Effects of antipsychotics and selective D3 antagonists on PPI deficits induced by PD 128907 and apomorphine. Behav Brain Res 2007;182:1-11.
- 29. Song SW, Sun Y, Su BL, Liu C, Yang C, Godfraind T, et al. Risperidone enhances the vulnerability to stroke in hypertensive rats. CNS Neurosci Ther 2012;18:343-9.
- 30. da Cruz Jung IE, Machado AK, da Cruz IB, Barbisan F, Azzolin VF, Duarte T, *et al.* Haloperidol and risperidone at high concentrations activate an *in vitro* inflammatory response of RAW 264.7 macrophage cells by induction of apoptosis and modification of cytokine levels. Psychopharmacology (Berl) 2016;233:1715-23.
- 31. Keilhoff G, Grecksch G, Bernstein HG, Roskoden T, Becker A. Risperidone and haloperidol promote survival of stem cells in the rat hippocampus. Eur Arch Psychiatry Clin Neurosci 2010;260:151-62.
- 32. Esch T, Stefano GB, Fricchione GL, Benson H. The role of stress in neurodegenerative diseases and mental disorders. Neuro Endocrinol Lett 2002;23:199-208.
- 33. Harrison PJ, McLaughlin D, Kerwin RW. Decreased hippocampal expression of a glutamate receptor gene in schizophrenia. Lancet 1991;337:450-2.
- 34. Radonjic NV, Knezevic ID, Vilimanovich U, Kravic-Stevovic T, Marina LV, Nikolic T, *et al.* Decreased glutathione levels and altered antioxidant defense in an animal model of schizophrenia: Long-term effects of perinatal phencyclidine administration. Neuropharmacology 2010;58:739-45.
- 35. Libman-Sokolowska M, Drozdowicz E, Nasierowski T. BDNF as a biomarker in the course and treatment of schizophrenia. Psychiatr Pol 2015;49:1149-58.
- 36. Li M, Du W, Shao F, Wang W. Cognitive dysfunction and epigenetic alterations of the BDNF gene are induced by social isolation during early adolescence. Behav Brain

- Res 2016;313:177-83.
- 37. Watanabe M, Fukuda A. Development and regulation of chloride homeostasis in the central nervous system. Front Cell Neurosci 2015;9:371.
- 38. Frost DO, Tamminga CA, Medoff DR, Caviness V, Innocenti G, Carpenter WT. Neuroplasticity and schizophrenia. Biol Psychiatry 2004;56:540-3.
- 39. Chen CC, Huang TL. Effects of antipsychotics on the serum BDNF levels in schizophrenia. Psychiatry Res 2011;189:327-30.
- Lee JG, Cho HY, Park SW, Seo MK, Kim YH. Effects of olanzapine on brain-derived neurotrophic factor gene promoter activity in SH-SY5Y neuroblastoma cells. Prog Neuropsychopharmacol Biol Psychiatry 2010;34:1001-6.
- 41. Chlan-Fourney J, Ashe P, Nylen K, Juorio AV, Li XM. Differential regulation of hippocampal BDNF mRNA by typical and atypical antipsychotic administration. Brain Res 2002;954:11-20.
- 42. Pillai A, Parikh V, Terry AV Jr., Mahadik SP. Long-term antipsychotic treatments and crossover studies in rats: Differential effects of typical and atypical agents on the expression of antioxidant enzymes and membrane lipid peroxidation in rat brain. J Psychiatr Res 2007;41:372-86.
- 43. Li M, He E, Volf N. Time course of the attenuation effect of repeated antipsychotic treatment on prepulse inhibition disruption induced by repeated phencyclidine treatment. Pharmacol Biochem Behav 2011;98:559-69.
- 44. Murray RM, Paparelli A, Morrison PD, Marconi A, Di Forti M. What can we learn about schizophrenia from studying the human model, drug-induced psychosis? Am J Med Genet B Neuropsychiatr Genet 2013;162B: 661-70
- Bernstein HG, Bogerts B, Keilhoff G. The many faces of nitric oxide in schizophrenia. A review. Schizophr Res 2005;78:69-86.

- 46. Pitsikas N. The role of nitric oxide donors in schizophrenia: Basic studies and clinical applications. Eur J Pharmacol 2015;766:106-13.
- 47. Pitsikas N. The role of nitric oxide synthase inhibitors in schizophrenia. Curr Med Chem 2016;23:2692-705.
- 48. Correll CU, Hauser M, Auther AM, Cornblatt BA. Research in people with psychosis risk syndrome: A review of the current evidence and future directions. J Child Psychol Psychiatry 2010;51:390-431.
- 49. Heinimaa M, Larsen TK. Psychosis: Conceptual and ethical aspects of early diagnosis and intervention. Curr Opin Psychiatry 2002;15:533-41.
- 50. McGlashan TH. Early detection and intervention in psychosis: An ethical paradigm shift. Br J Psychiatry Suppl 2005;48:s113-5.
- 51. Weiss MG, Ramakrishna J, Somma D. Health-related stigma: Rethinking concepts and interventions. Psychol Health Med 2006;11:277-87.
- 52. Corson PW, Nopoulos P, Miller DD, Arndt S, Andreasen NC. Change in basal ganglia volume over 2 years in patients with schizophrenia: Typical versus atypical neuroleptics. Am J Psychiatry 1999;156:1200-4.
- 53. DeLisi LE. The concept of progressive brain change in schizophrenia: Implications for understanding schizophrenia. Schizophr Bull 2008;34:312-21.
- 54. Filakovic P, Degmecic D, Koic E, Benic D. Ethics of the early intervention in the treatment of schizophrenia. Psychiatr Danub 2007;19:209-15.
- 55. Tan QR, Wang XZ, Wang CY, Liu XJ, Chen YC, Wang HH, *et al.* Differential effects of classical and atypical antipsychotic drugs on rotenone-induced neurotoxicity in PC12 cells. Eur Neuropsychopharmacol 2007;17:768-73.
- Liu YP, Yang YY, Wan FJ, Tung CS. Importance of intervention timing in the effectiveness of antipsychotics. Prog Neuropsychopharmacol Biol Psychiatry 2017. pii: S0278-5846(16)30121-X.